# ab208572 - Universal HIER antigen retrieval reagent (10X)

For heat-induced epitope retrieval of formalin-fixed paraffin-embedded tissues For research use only - not intended for diagnostic use.

For overview, typical data and additional information please visit: <a href="www.abcam.com/ab208572">www.abcam.com/ab208572</a> (use <a href="www.abcam.co.jp/ab208572">www.abcam.co.jp/ab208572</a> for China, or <a href="www.abcam.co.jp/ab208572">www.abcam.co.jp/ab208572</a> for Japan)

#### **Precautions**

## Please read these instructions carefully prior to beginning the assay.

- We understand that, occasionally, experimental protocols might need to be modified to meet unique experimental circumstances. However, we cannot guarantee the performance of the product outside the conditions detailed in this protocol booklet.
- Please review the Safety Datasheet (SDS) provided with the product for information on how to handle and dispose of this reagent.
- Observe good laboratory practices. Gloves, lab coat, and protective eyewear should always be worn. Never pipet by mouth. Do not eat, drink or smoke in the laboratory areas.
- All biological materials should be treated as potentially hazardous and handled as such. They should be disposed of in accordance with established safety procedures.
- Avoid contacting the skin and mucous membranes with reagents and specimens. If reagents or specimens come in contact with sensitive areas, wash with copious amounts of water.

# **Materials Supplied**

Item	Quantity	Storage temperature
Universal HIER antigen retrieval reagent (10X)	100 mL	RT

Storage and Stability: Store product at room temperature. See product label for expiry date.

#### Materials Required, Not Supplied

These materials are not included, but will be required to successfully perform this assay:

- Deionised water
- Pipettes and pipette tips, including multi-channel pipette
- Assorted glassware for the preparation of reagents and buffer solutions
- Microscope slides, positively charged
- Drying oven
- Positive and negative tissue controls
- Xylene (or xylene substitute)
- Ethanol or reagent alcohol
- Pressure cooker
- Deionized or distilled water
- Wash buffer (TBS/PBS)
- Enzymes for digestion

- Avidin-Biotin Blocking Kit (if using Labeled Streptavidin-Biotin (LSAB) or avidin-biotin complex kits for detection)
- Peroxidase block
- Protein block or suitable serum
- Primary antibody
- Negative control reagents
- Detection kits
- Chromogens
- Hematoxylin
- Bluing reagent
- Mounting medium

#### Assay procedure

Equilibrate all materials and prepared reagents to correct temperature prior to use.

- Deparaffinize tissues and hydrate with water. If necessary, block for endogenous peroxidase and wash in deionised water.
- Dilute concentrated Universal HIER antigen retrieval reagent (10X) at a ratio of 1:10 (1 mL Universal HIER antigen retrieval reagent (10X) to 9 ml of deionized water).
- Place slides into 1X retrieval solution in a slide container (e.g. Coplin Jar, staining dish or metal slide canister).
- Retrieve sections using heat. Follow the recommendations on the antibody data sheet and/or refer to Abcam's IHC antigen retrieval protocol (www.abcam.com/protocols/ihcantigen-retrieval-protocol).
- Check solution for appropriate color change. (See Technical Notes)
- Gently rinse by gradually adding DI water to the solution, then remove slides and rinse with DI water.

#### **Technical Notes**

- Concentrated Universal HIER antigen retrieval reagent (10X) is a bright yellow color. The RTU or 1X solution is a pale-yellow color. When the solution reaches 80-125°C, the solution turns yellow and indicates that the high temperature solution is at correct pH. Should the pH rise above 7.0, the solution turns a fuschia red color. Should the pH drop too low, the solution turns a pink color.
- Use positive charged slides and cut tissues into 4-5-micron sections. Do not use any
  adhesives in the water bath. Poor fixation and processing of tissues will cause tissue
  sections, especially fatty tissues such as breast, to fall off the slides. Tissues should be fixed
  for a minimum of 6-12 hours.
- Protocol time and temperatures for HIER can vary depending on the heat source used.

### **Technical Support**

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